

Genetic Techniques in the Evaluation of Short Stature

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KEYWORDS

- Short stature • Height • Genome-wide analysis • Microarray analysis
- Whole-exome sequencing

KEY POINTS

- Many children with short stature do not have a cause to explain the disorder.
- Several genetic techniques, some of which are clinically available and others at the research stage, are helping to identify potential variants to explain short stature.
- Understanding these techniques will assist clinicians in their appropriate use and also help to foster support in expanding genetic screening to delineate mechanisms of poor growth.

INTRODUCTION

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Normal growth is a complex dynamic process dependent on the coordination of multiple factors, including genetics and nutrition, as well as biological factors such as hormones, all working in balance. Any acute or chronic pathologic process may interfere with normal growth.¹ The typical endocrine evaluation comprises a history, physical examination, and laboratory testing, including hormone levels and radiological studies. A genetic evaluation usually depends on initial clinical and laboratory findings. In many cases an endocrine, nutritional, or chromosomal abnormality is not apparent. However, the expansion of genetic technology has increased the diagnostic yield of genetic testing. Ongoing research efforts to identify genes influencing growth will provide a better understanding of mechanisms underlying abnormal growth and will eventually lead to novel management approaches.

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49 This article focuses on describing some of the current advances in genetic testing
50 that can be used as part of the diagnostic evaluation of children with short stature.
51 This article is not intended to be comprehensive, but rather to educate the reader
52 on the increasing efficiency of genetic screening tools that will eventually provide cli-
53 nicians with important diagnostic capabilities for the evaluation of children with
54 abnormal stature.

55 56 GENETIC TESTING STRATEGIES

57 The approach to understanding disorders at the level of the DNA currently has several
58 options. Depending on the clinical phenotype or the specific trait of interest, clinicians
59 or investigators have several choices for how to screen a patient or cohort of patients.
60 Some testing is commercially available, whereas other genetic tests have not been
61 fully validated or standardized and are only available through research protocols.
62 This testing ranges from conventional chromosome analysis and microarray analysis,
63 to sequencing of single genes, sequencing of a panel of genes, or to whole-exome
64 sequencing (WES). Each technique has advantages as well as limitations. The article
65 explores how these techniques have affected clinical practice and helped further the
66 understanding of the mechanisms of growth. Abnormal expression of imprinted genes
67 has been associated with overgrowth and poor growth in syndromes such as
68 Beckwith-Wiedemann and Russell-Silver syndrome. Methylation testing to detect
69 active and inactive genes in imprinted areas of the genetic material is also available
70 to screen children presenting with features suspicious for these syndromes. However,
71 this group of disorders is not discussed in detail.

72 73 74 CHROMOSOME ANALYSIS

75 Chromosomal analysis or karyotyping has become an effective and efficient tool in the
76 diagnosis of patients with short stature. It involves the visual examination of G-banded
77 chromosomes in order to detect large structural deletions and duplications, mono-
78 somies, trisomies, and balanced rearrangements. Typically, these patients present
79 with characteristic phenotypic features that include short stature. An example is
80 mosaic Turner syndrome; however, short stature in a female patient may be sufficient
81 justification to study chromosomes and thus has been argued as means to identify
82 Turner syndrome at an earlier age.² In the case of tall stature, phenotypic features
83 of Klinefelter syndrome (XXY), is another example in which the evaluation of chromo-
84 somes is useful. Fluorescence in situ hybridization (FISH) can complement karyotyp-
85 ing in order to detect various small chromosomal abnormalities, such as deletions and
86 duplications. This technique has been used, for example, to detect whole-gene dele-
87 tions of the short stature homeobox-containing gene (*SHOX*), which have been asso-
88 ciated with Leri-Weill dyschondrosteosis (LWD), but have also been implicated as a
89 cause of short stature in children in whom full endocrine evaluations have not identified
90 causes for their poor growth.³

91 A karyotype with FISH may also detect specific suspected microdeletion syn-
92 dromes. Deletions of chromosome 22q11.2 are among the commonest of all microdele-
93 tions and are associated with short stature, often in conjunction with other birth
94 defects, typically congenital heart defects or cleft palate (DiGeorge or velocardiofacial
95 syndromes). However, the presence of a supernumerary chromosome derived from
96 chromosome 22 has been reported to cause cat-eye syndrome.^{4,5} It is associated
97 with anal atresia, coloboma of the iris, ear malformations, and short stature, and in
98 rare cases growth hormone (GH) deficiency.⁵ In another example, there is a

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100 description of a patient with an Xp13.2 chromosome microduplication, who presented
101 with features similar to Turner syndrome that included short stature.⁶

102 Karyotyping and FISH are easily accessible to clinicians and economical, but have
103 been low yield in cases of short stature without a cause. Most children with chromo-
104 somal anomalies and microdeletion/duplications have dysmorphic features, congeni-
105 tal anomalies, and intellectual disability, and treatment of short stature would be of
106 limited utility. Nevertheless, the recognition of phenotypic features would be important
107 in determining priorities for care and would alert clinicians to the risk of associated
108 comorbidities. In isolated short stature, a chromosomal cause is unlikely. For example,
109 identifiable gene deletions in the SHOX gene for patients who present with short stat-
110 ure reportedly only yields a diagnosis in 2% to 15% of this patient population.⁷

112 TARGETED OR MULTIGENE SEQUENCING

113 Phenotypic or hormonal abnormalities may prompt the investigation of a specific gene
114 abnormality. For example, a suspicion of a skeletal dysplasia because of disproport-
115 ionate short stature may prompt testing of the *FGFR3* gene. Suspicion of a rasopathy
116 such as Noonan syndrome can prompt specific testing for *PTPN11* and related genes
117 in the ras-MAPK pathway. Commercial laboratories now offer multigene panels for
118 testing of proportionate and disproportion short stature. This approach has led
119 to identification of mutations in genes that may not be suspected on the clinical eval-
120 uation. The biggest limitation of using multigene panel testing and newer next-
121 generation sequencing (NGS) technology to screen large numbers of genes is the
122 identification of sequence variants that are difficult to interpret (variants of unknown
123 significance).
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125 In patients with short stature, much focus is placed on identifying abnormalities
126 within the hypothalamic-pituitary-GH axis. These studies typically use a targeted
127 gene approach, which may include screening for mutations in *GH1*, GHRH receptor
128 (*GHRHr*) or insulinlike growth factor 1 receptor (*IGF-1R*). Although researchers have
129 been able to identify genetic perturbations leading to abnormalities in hormone secre-
130 tion, hormone viability, or receptor sensitivity, those patients who have identified ge-
131 netic causes in this axis continue to be rare. Even in those patients with identified
132 genetic abnormalities, there is often no clear genotype-phenotype correlation among
133 patients with similar mutations.⁸ Therefore clinical testing for specific genes involved in
134 the GH axis has limited utility at this time.^{9,10} A recent report characterizing the lack of
135 differences in the auxologic and biochemical parameters of short children with *SHOX*
136 variants compared with children with normal *SHOX* also helps to make this point.¹¹

138 GROWTH HORMONE DEFICIENCY

139 Isolated GH deficiency (IGHD) is a rare diagnosis with several described mutations in
140 both the *GH1* and the *GHRHr* gene, but no cases of *GHRH* gene mutation have been
141 recorded. Less than 15% of patients with IGHD have an identified gene defect,
142 although the rate increases up to almost 35% with investigations of familial cases.¹²
143 A recent report describes an autosomal form of IGHD over 3 generations secondary
144 to a heterozygous missense mutation in the *GH-1* gene leading to misfolding of the
145 protein.¹³ This type of IGHD is known as type II GH deficiency (GHD) and the severity
146 of the phenotype has been correlated with the type of mutation identified in pa-
147 tients.^{14,15} In these cases in which GHD is thought to be secondary to mutations
148 affecting splicing of *GH1*, there is production of deleterious isoforms leading to a
149 dominant negative effect on the secretion of the 22-kDa full-length peptide.^{15,16} The
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151 height deficit in these patients is variable and there is also a risk for developing addi-
152 tional pituitary hormone deficiencies.

153 Recently, a heterozygous mutation in the *POU1F1* gene (p.Pro76Leu), which maps
154 to a conserved site within the transactivation domain, was described as a cause of
155 IGHD. Classically, mutations in the *POU1F1* gene lead to a clinical phenotype triad
156 of somatotroph, lactotroph, and thyrotroph dysfunction. Functional studies of this mu-
157 tation show how this mutation can alter specific transactivating functions as well as
158 interactions with specific cofactors such as ELK1, which activates endogenous
159 GH.¹⁷ These studies report not only a novel mutation in *POU1F1* leading to IGHD
160 but further elucidate the complexities of pituitary development and function.

162 MULTIPLE PITUITARY HORMONE DEFICIENCIES

163 Along the spectrum of defects along the GHRH-GH axis, results of ongoing research
164 continue to help clinicians better understand the development of multiple pituitary hor-
165 mone deficiencies (MPHDs), of which abnormal stature is typically a part. Neverthe-
166 less, this field continues to be complicated because clinical phenotypes are
167 typically not well predicted by genotypes.^{18,19} MPHD is often a dynamic progression
168 of disorder because patients may develop additional hormone deficiencies over time.
169 Although it is clear that idiopathic MPHD results from aberrations in pituitary develop-
170 ment that may affect structure, cellular function, or even cell survival, most of these
171 disorders do not have an identifying mutated gene and may suggest that MPHD is
172 most likely not a single-gene abnormality. Despite this idea, several mutated genes
173 have been reported as likely causes of the development of hypothalamic and pituitary
174 disease.^{8,20,21} A better understanding of pituitary development and its complex regu-
175 lation will most likely help to provide insight into potential pathologic mechanisms
176 leading to hormone dysfunction.²² There is still much unknown about the coordination
177 of the events, including expression of transcription factors and signaling genes,
178 required for a process that is temporally and spatially dependent in order to produce
179 a functional gland. **Table 1** summarizes several of the reported genes shown to cause
180 short stature and, in most cases, other endocrine abnormalities.

182 TARGETING BONE GROWTH

183 In bone growth, both C-type natriuretic peptide (CNP) and its receptor, natriuretic pep-
184 tide receptor-B (NPR-B), have been cited as important regulators of endochondral
185 bone growth.²³ Studies using mouse models in which there is a loss-of-function mu-
186 tation of the *Npr2* gene or loss of CNP show that these conditions result in disproportion-
187 ate dwarfism and abnormal endochondral ossification.^{24,25} In addition, disruption Q15
188 of the CNP-mediated signaling via cGMP in both in vivo and in vitro studies leads to
189 skeletal defects at the growth plate.²⁶ The importance of these factors in humans
190 has also been recognized in studies of normal and abnormal growth.^{27,28}

191 Homozygous mutations in *NPR2* are associated with a severe dysplasia known as
192 acromesomelic dysplasia, type Maroteaux (AMDM; OMIM [Online Mendelian Inheri- Q16
193 tance in Man] #602875).²⁹ These patients have severe short stature, shortening of
194 extremities, and bowing of the forearm, which clinically is similar to LWD (OMIM
195 #127300). Despite this extreme phenotype with homozygous mutations on this
196 gene, some investigators have described heterozygous mutations of *NPR2* as a
197 potential cause of disorder in patients otherwise classified as ISS.³⁰ In one small study Q17
198 of 47 patients diagnosed with ISS, the investigators reported that 6% of the patients
199 had heterozygous *NPR2* mutations.³⁰ In a larger group of patients with short stature,
200 only 2% of the patients had identified heterozygous *NPR2* mutations.³¹ Given the
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Table 1
Single-gene candidate analysis

Disorder	Subtypes	Gene Mutation	Description
IGHD	IGHD 1A	<i>GH</i> (gross deletion, microdeletion)	Absent GH, severe short stature, development of anti-GH Abs after treatment with GH, tx with IGF-1
	IGHD 1B	<i>GH</i> (splice-site mutations)	Short stature, tx with rGH
	IGHD II	<i>GH</i> (splice-site mutations, intron/exon splice enhancer, missense mutations)	Short stature, tx with rGH
			Short stature, agammaglobulinemia, X-linked
Multiple pituitary hormone deficiency	—	<i>HESX1</i>	Associated with septo-optic dysplasia or MPHD. Deficiencies: GH, TSH, gonadotropins, evolving ACTH
	—	<i>PROP1</i>	Deficiencies: GH, TSH, Prl, possibly gonadotropins, ACTH; associated hypoplastic or enlarge pituitary
	—	<i>POU1F1 (PIT1)</i>	Deficiencies: GH, TSH, Prl; hypoplastic pituitary
	—	<i>LHX3</i>	Deficiencies: GH, PRL, LH, FSH, and TSH; associated with rigid cervical spine/limited head rotation
	—	<i>LHX4</i>	Deficiencies: GH, variable TSH, LH, FSH, ACTH; possible a target of POU1F1
	—	<i>OTX2</i>	Deficiencies: GH, TSH, LH, FSH, ACTH; associated with eye abnormalities
	—	<i>SOX2</i>	Deficiencies: LH, FSH, variable GHD; anophthalmia/microphthalmia, hypothalamic hamartoma
—	<i>GLI2</i>	Deficiencies: GH, TSH, LH, FSH, ACTH; holoprosencephaly, craniofacial abnormalities	

Abbreviations: Abs, antibodies; ACTH, adrenocorticotrophic hormone; FSH, follicle-stimulating hormone; LH, luteinizing hormone; Prl, prolactin; rGH, recombinant growth hormone; TSH, thyroid-stimulating hormone; tx, treatment.

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clinical similarities of AMDM and LWD and that in cases of LWD about 30% of patients do not have identified *SHOX* mutations, other investigators have screened patients with LWD for mutation in *NPR2*. Only 3% of patients in a pool of 173 had *NPR2* mutations, which included a total of 7 variants; however, functional studies confirmed that 6 of the *NPR2* variants were pathogenic mutations.³²

Despite these low percentages, it needs to be considered that these studies looked at small numbers of patients and, in addition, the phenotypic variability seen in short stature may account for a seemingly limited role of *NPR2*. An earlier study predicted that up to 1 in 30 individuals with idiopathic short stature (ISS) are carriers of *NPR2* mutations.³³ More recently, another study of patients with both short stature and normal stature provided further evidence of the impact of heterozygous mutations in *NPR2* and concluded that patients with ISS should be screened.³⁴ The investigators reported that *NPR2* loss-of-function variants may account for 0.4% to 4% of patients with short stature and is associated with mild and variable growth impairment without a distinct skeletal

phenotype. These conclusions are further supported by the finding that, when familial cases were analyzed, 13.6% of cases had *NPR2* loss-of-function variants.

In contrast with these studies, the importance of *CNP* in the regulation of human endochondral bone growth has also been suggested in studies of patients with bone overgrowth.³⁵ Further support for the importance of *NPR2* is shown in a report of a family with tall stature and macrodactyly of both great toes; all were found to harbor a gain-of-function type mutation in the *NPR2* gene.³⁶

GENOMIC TECHNOLOGIES

The ability to scan the human genome has provided science with powerful tools to identify novel genes and study their role in normal physiology. Because normal variation in height is not only a heritable trait but also a polygenic trait, the advantages of genome-wide association studies (GWAS) are clear.^{37–39} It has been reported that GWAS have been able to identify hundreds of variants associated with human traits, but may not necessarily account for phenotypic variation. An earlier report using a genome-wide scan reported 20 variants associated with adult height, although these accounted for only approximately 3% of the height variation in the studied population.⁴⁰ Genome-wide analysis studies have also helped to identify at least 180 loci thought to have an influence on height.⁴¹ These loci were not random, but were statistically associated with genes that underlie skeletal growth defects.⁴¹

Given the vast amount of information provided by analyzing the whole genome, strategies can be designed to only study a more select group of genes, such as in a microarray, or conduct more global screening, such as is seen with WES or even whole-genome sequencing. Each of these strategies provides advantages as well as disadvantages, but nevertheless has continued to improve understanding of the complexities of stature.

CHROMOSOME MICROARRAY ANALYSIS

Chromosome microarray or array Comparative genomic hybridization (CGH) is an approach that uses hybridization of a patient's DNA and reference DNA to known DNA sequences, which can be oligonucleotides or single nucleotide polymorphism (SNP), from across the genome, in order to detect submicroscopic deletions and duplications. This technology can also detect uniparental disomy by detecting areas of SNP homozygosity. Copy number variant (CNV) is a term that refers to either heritable or novel variations in the number of gene copies. These duplications or deletions, which can include 1 or several genes, can be benign genetic variations in humans or a cause of pathologic phenotypes.⁴² This condition has been investigated as mechanism of disorder in other phenotypes with variable yield.^{43,44}

Some groups have reported CNVs in up to 10% of populations with short stature and height 2 standard deviation (SD) scores less than the average with no underlying cause to explain stature.^{45,46} Although the traditional candidate gene approach has allowed clinicians to diagnose monogenic defects leading to short stature, some believe that studying patients with extremes in stature (short or tall) in order to identify CNVs could potentially identify CNVs that include novel genes leading to short stature.⁴⁶

In one study of 162 patients with short stature, the investigators identified 40 potentially pathogenic CNVs in 33 families; however, further investigation and segregation analysis of these identified genes, as well as the previous documented literature, led to the conclusion that only 4% of these CNVs were confidently associated with short stature.⁴⁶ One recent report found that CNV analysis in a family with short

304 stature revealed that the affected patient carried a paternally inherited heterozygous o20
305 IGF-1 gene deletion, thus suggesting that CNVs may play a role in disrupting the
306 GH-IGF-1 axis.⁴⁷ Despite seemingly low yield in identifying target genes, earlier
307 studies suggest that patients with short stature carry a greater global burden of
308 CNVs. Nevertheless, these seem to be lower frequency CNVs and in the context of
309 a polygenic trait such as stature, how much CNVs contribute to normal physiology
310 versus disorder continues to be under investigation.⁴⁸

311 Array-based comparative genomic hybridization/chromosome microarray analysis
312 has been used in the study of children born small for gestational age (SGA) who
313 have failure of catch-up growth. Some clinicians have used a candidate gene
314 approach (*SHOX*, *IGF-1*, and *IGF-1R*) to identify a genetic cause, but this is reported
315 to account for approximately 6% of these short children.⁴⁹ However, array-based
316 genomic copy number analysis found that 16% of this population may carry rare path-
317 ogenic or probably pathogenic CNVs.⁵⁰ In another study, 27% of SGA patients studied
318 with whole-genome SNP array analysis had identified CNVs that could potentially
319 explain the short stature.⁵¹ The CNVs that were considered potentially pathogenic
320 included several potential candidate genes previously implicated in GWAS studies
321 of height or skeletal disorders in rodents.^{41,51}

322 Because prenatal and postnatal growth in SGA children is polygenic and influenced
323 by maternal, placental, and fetal factors, this could be considered as a cohort of
324 patients who could provide better insight into understanding genetic determinants
325 of growth and development. However, some clinicians think that gene variants only
326 account for a small fraction of genetic variation and genome scans may be more help-
327 ful for focusing on rarer variants to gather insight into the significant pathologic
328 mechanisms.⁵²

330 WHOLE-GENOME AND WHOLE-EXOME SEQUENCING

332 The technology referred to as NGS, which is also known as high-throughput
333 sequencing, has made it possible to screen large numbers of genes rapidly. It can pro-
334 vide higher efficiency in identifying pathologic genes and is less labor intensive than
335 selective gene analysis. Furthermore, given the limitations of current diagnostic mo-
336 dalities in endocrinology and conditions in which disorders may evolve over time,
337 NGS is a possible tool to identify novel genes. The limitation of this technology is in
338 interpretation of sequence variants detected by analysis. Multiple tools are used to
339 help interpretation, including population frequencies, disease frequencies, in silico
340 protein prediction, family segregation, and algorithms that have been devised to stan-
341 dardize its interpretation. This technology would be more useful for identifying rare
342 variants versus more common variants.⁵²

343 One study screened patients with short stature for more than 1000 genes, many of
344 which have been associated with growth disorders, skeletal dysplasia, growth plate
345 biology, or GH signaling, and reported 1349 novel variants. The investigators high-
346 lighted probably pathogenic variants in IGF-1R and known pathogenic variants in
347 the *PTPN11* gene, which is associated with Noonan syndrome.⁵³ In a larger study,
348 the investigators reported almost 700 variants in 423 loci in tissue types and pathways
349 involved in growth.⁵⁴ These variants were reported to account for 36% of the herita-
350 bility of adult height.

351 An important application of NGS is in the development of WES, which is a high-
352 throughput genetic analysis in which the coding DNA of patients is sequenced.
353 Although the exome is about 1% of the entire genome, it provides a more efficient
354 method of analysis and limits the massive amounts of data screened. It has been

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Table 2
Genetic technology used in the study of short stature

Technique	Description	Indications	Advantages	Limitations	Other	
Chromosome analysis	Karyotype	Microscopic G-banded chromosomes	Short stature associated with syndromic features, multiple anomalies, developmental delay	Economical Easily accessible Provides definitive diagnosis of monosomies, trisomies, and large deletions/duplications	Limited to syndromic short stature with low yield because of limited resolution	Used in combination with other cytogenetic techniques such as FISH (eg, diagnosis of <i>SHOX</i> haploinsufficiency)
	Microarray analysis	Comparative hybridization of patient and reference DNA to oligonucleotides and SNPs from across the genome	Similar to chromosome analysis	Efficient screen for copy number variation. Can detect uniparental disomy. May replace chromosome analysis because of higher resolution and detection of submicroscopic as well as large abnormalities	Detects variants of unknown significance that may be difficult to interpret	—
Molecular diagnostics	Single-gene analysis	Targeted screening of specific genes using Sanger sequencing	Short stature disorder suspected by clinical features, monogenic disorder	Detects many types of mutations	Targeted to specific genes only. Time consuming and expensive	Commonly used to screen for specific genes causing disorders of hypothalamic-pituitary-GH axis

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	Gene panel testing	Next-generation sequencing for screening of multiple genes that may be causing the phenotype	Genetically heterogeneous disorders and overlapping phenotypes	Rapid and less expensive, can screen large numbers of genes efficiently	Needs Sanger confirmation May identify variants of unknown significance. Cannot detect certain types of mutations	Genes can be selected based on phenotype to evaluate for isolated or syndromic short stature
	WES	Analysis of exons of the entire genome using high-throughput DNA sequencing technology	Genetic short stature, but no known single-gene defect is detected with other techniques, or not known	Identify cause of uncommon or rare conditions Efficient screening; targets only protein coding (exons) Efficient tool to identify pathologic variant in conditions with familial inheritance	Identifies variants of unknown significance. Cannot detect certain types of mutations Variants in genes with unknown function may be detected. Incidental findings may be discovered in genes unrelated to phenotype Large data sets require ongoing standardization/variant analyses remains a challenge	—

457 estimated, based on the rate of people having genetic disorders, that the molecular
458 diagnostic yield of WES is up to 25% compared with using a single-gene approach,
459 which yielded results in less than 15% of patients.⁵⁵ Other investigators who have
460 used WES on adult patients have reported similar results, with a diagnostic rate of
461 17.5%.⁵⁶ Reports have commented that the utility of this technology has helped to
462 produce unexpected findings of de novo mutations that would not have been identi-
463 fied with more traditional genetic screenings.

464 In endocrinology, there has been some application of WES in the study of disorders
465 such as type 2 diabetes mellitus, hypogonadism, and endocrine neoplasia.⁵⁷ Muta-
466 tions within individual genes may play a major role in determining an individual's height
467 and, with the assumption that unidentified highly penetrant genetic variants contribute
468 to stature, WES will continue to be an effective tool in advancing the field. In one recent
469 report, WES was performed on patients identified with severe short stature more than 021
470 3 SD less than the mean for age, and the investigators reported that, in 36% of the
471 families analyzed, a causal gene was identified.⁵⁸ As with other forms of genetic anal-
472 ysis, the strategy of screening cohorts of families to help filter out normal variants may
473 help to better target more pathogenic variants.

474 One example of how WES has provided helpful insight into short stature is a recent
475 report that identified a gene that encodes aggrecan, a proteoglycan in the extracellular
476 matrix of growth plate and other cartilaginous tissues, known as *ACAN*. A heterozy-
477 gous gene mutation was identified in individuals with short stature along with an
478 advanced bone age through WES and predicted to adversely affect protein function.⁵⁹
479 The described heterozygous mutations in *ACAN* seem to lead to mild skeletal
480 dysplasia.

481 As with other methods of genomic analysis, the limitations of WES include the
482 inability to recognize several potential genetic pathogenic causes of disease, including
483 important intronic regions or splice-site sequences, structural variants of DNA such as
484 translocations and inversions, genomic imprinting, epigenetic changes, and CNVs that
485 do not change DNA sequence. Furthermore, another limitation may be in the failure to
486 efficiently identify susceptibility genes in common disease traits, such as short stature,
487 that are complex and multifactorial. Susceptibility traits with generally low penetrance
488 and additive effects rarely are a primary cause of disorder. However, the continued
489 use of WES in clinical practice will inevitably provide better insights into rare,
490 higher-penetrance monogenic causes of short stature.

491 SUMMARY

492 It has been reported in a recent study that the standard medical evaluation for short
493 stature only had a 1.3% diagnostic yield for patients and thus shows the need to
494 expand diagnostic modalities for clinicians.⁹ The advances in understanding genetic
495 determinants of height will increase as researchers focus on specific phenotypes,
496 which could include the presentations of severe familial short stature, short stature
497 associated with specific syndromes, or children at risk for poor growth such as
498 SGA. Genetic tools and analyses are quickly revolutionizing the approach to under-
499 standing polygenic traits such as stature and will continue to assist in the identification
500 of monogenic disorders leading to abnormal stature.⁶⁰ This research will inevitably
501 help to expand on the current limited therapies to treat short stature.⁶¹ **Table 2** sum-
502 marizes several genetic techniques reviewed in this article, summarizing indications
503 and advantages for using each technique in the evaluation of stature. Such indications
504 and limitations apply to other areas of medicine as well.^{62,63} The endocrine evaluation
505 for stature will notably continue to evolve as technologies for genetic screening
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508 improve. Each of the described techniques in this article has limitations in its ability to
509 specifically target the causes of short stature, but each has helped to expand the
510 appreciation of the complexities of height determination. The accessibility of patients
511 to participate in research studies will increase and science will delineate the mecha-
512 nisms of how genes determine stature. Once the mechanisms of disorders are better
513 outlined, science will then be able to provide better diagnostic tools, along with novel
514 therapeutic options, to address abnormal stature. Genetic screening options with
515 better-defined algorithms will ultimately become part of a new standard approach
516 for clinicians treating patients with short stature.

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